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(54) Title: NOVEL HUMAN TRANSPORTER PROTEINS AND POLYNUCLEOTIDES ENCODING THE SAME

(57) Abstract: Novel human polynucleotide and polypeptide sequences are disclosed that can be used in therapeutic, diagnostic, and pharmacogenomic applications.

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NOVEL HUMAN TRANSPORTER PROTEINS AND
POLYNUCLEOTIDES ENCODING THE SAME

5 The present application claims the benefit of U.S. Provisional Application Number 60/210,045 which was filed on June 7, 2000 and is herein incorporated by reference in its entirety.

1. INTRODUCTION

10 The present invention relates to the discovery, identification, and characterization of novel human polynucleotides encoding proteins that share sequence similarity with mammalian transporter proteins. The invention encompasses the described polynucleotides, host
15 cell expression systems, the encoded proteins, fusion proteins, polypeptides and peptides, antibodies to the encoded proteins and peptides, and genetically engineered animals that either lack or over express the disclosed polynucleotide sequences, antagonists and agonists of the
20 proteins, and other compounds that modulate the expression or activity of the proteins encoded by the disclosed polynucleotide sequences that can be used for diagnosis, drug screening, clinical trial monitoring, the treatment of diseases and disorders, and cosmetic or nutraceutical
25 applications.

2. BACKGROUND OF THE INVENTION

Transporter proteins are integral membrane proteins that mediate or facilitate the passage of materials across the lipid bilayer. Given that the transport of materials
30 across the membrane can play an important physiological role, transporter proteins are good drug targets. Additionally, one of the mechanisms of drug resistance involves diseased cells using cellular transporter systems to export

chemotherapeutic agents from the cell. Such mechanisms are particularly relevant to cells manifesting resistance to a multiplicity of drugs.

3. SUMMARY OF THE INVENTION

5 The present invention relates to the discovery, identification, and characterization of nucleotides that encode novel human proteins; and the corresponding amino acid sequences of these proteins. The novel human proteins (NHPs) described for the first time herein share structural
10 similarity with mammalian ion transporters, sulfate transporters, and particularly the sulfate transporter that has been associated with diastrophic dysplasia.

 The novel human nucleic acid sequences described herein, encode alternative proteins/open reading frames
15 (ORFs) of 679, 621, 663, 605, 656, and 598 amino acids in length (see respectively SEQ ID NOS: 2, 4, 6, 8, 10, and 12. The invention also encompasses agonists and antagonists of the described NHPs, including small molecules, large molecules, mutant NHPs, or portions thereof, that compete
20 with native NHP, peptides, and antibodies, as well as nucleotide sequences that can be used to inhibit the expression of the described NHPs (e.g., antisense and ribozyme molecules, and gene or regulatory sequence replacement constructs) or to enhance the expression of the
25 described NHP polynucleotides (e.g., expression constructs that place the described polynucleotide under the control of a strong promoter system), and transgenic animals that express a NHP transgene, or "knock-outs" (which can be conditional) that do not express a functional NHP. Knock-out
30 mice can be produced in several ways, one of which involves the use of mouse embryonic stem cells ("ES cells") lines that contain gene trap mutations in a murine homolog of at least

one of the described NHPs. When the unique NHP sequences described in SEQ ID NOS:1-13 are "knocked-out" they provide a method of identifying phenotypic expression of the particular gene as well as a method of assigning function to previously
5 unknown genes. Additionally, the unique NHP sequences described in SEQ ID NOS:1-13 are useful for the identification of coding sequence and the mapping a unique gene to a particular chromosome.

Further, the present invention also relates to
10 processes for identifying compounds that modulate, *i.e.*, act as agonists or antagonists, of NHP expression and/or NHP activity that utilize purified preparations of the described NHPs and/or NHP product, or cells expressing the same. Such compounds can be used as therapeutic agents for the treatment
15 of any of a wide variety of symptoms associated with biological disorders or imbalances.

4. DESCRIPTION OF THE SEQUENCE LISTING AND FIGURES

The Sequence Listing provides the sequences of NHP ORFs
20 encoding the described NHP amino acid sequences. SEQ ID NO:13 shows a NHP ORF and flanking regions.

5. DETAILED DESCRIPTION OF THE INVENTION

The NHPs described for the first time herein are novel
25 proteins that may be expressed in, *inter alia*, human cell lines, pituitary, lymph node, kidney, testis, thyroid, heart, placenta, adipose, placenta, trachea, umbilical vein endothelium, fetal brain, and fetal kidney cells.

The present invention encompasses the nucleotides
30 presented in the Sequence Listing, host cells expressing such nucleotides, the expression products of such nucleotides, and: (a) nucleotides that encode mammalian homologs of the described polynucleotide sequences, including the

specifically described NHPs, and the NHP products;

(b) nucleotides that encode one or more portions of the NHPs that correspond to functional domains, and the polypeptide products specified by such nucleotide sequences, including

5 but not limited to the novel regions of any active domain(s);

(c) isolated nucleotides that encode mutant versions, engineered or naturally occurring, of the described NHPs in which all or a part of at least one domain is deleted or altered, and the polypeptide products specified by such

10 nucleotide sequences, including but not limited to soluble proteins and peptides in which all or a portion of the signal (or hydrophobic transmembrane) sequence is deleted; (d) nucleotides that encode chimeric fusion proteins containing all or a portion of a coding region of an NHP, or one of its

15 domains (e.g., a receptor or ligand binding domain, accessory protein/self-association domain, etc.) fused to another peptide or polypeptide; or (e) therapeutic or diagnostic derivatives of the described polynucleotides such as oligonucleotides, antisense polynucleotides, ribozymes,

20 dsRNA, or gene therapy constructs comprising a sequence first disclosed in the Sequence Listing.

As discussed above, the present invention includes:

(a) the human DNA sequences presented in the Sequence Listing (and vectors comprising the same) and additionally

25 contemplates any nucleotide sequence encoding a contiguous NHP open reading frame (ORF) that hybridizes to a complement of a DNA sequence presented in the Sequence Listing under highly stringent conditions, e.g., hybridization to filter-bound DNA in 0.5 M NaHPO₄, 7% sodium dodecyl sulfate (SDS),

30 1 mM EDTA at 65°C, and washing in 0.1xSSC/0.1% SDS at 68°C (Ausubel F.M. et al., eds., 1989, Current Protocols in Molecular Biology, Vol. I, Green Publishing Associates, Inc., and John Wiley & sons, Inc., New York, at p. 2.10.3) and

encodes a functionally equivalent gene product. Additionally contemplated are any nucleotide sequences that hybridize to the complement of a DNA sequence that encodes and expresses an amino acid sequence presented in the Sequence Listing
5 under moderately stringent conditions, e.g., washing in 0.2xSSC/0.1% SDS at 42°C (Ausubel et al., 1989, *supra*), yet still encodes a functionally equivalent NHP product. Functional equivalents of a NHP include naturally occurring NHPs present in other species and mutant NHPs whether
10 naturally occurring or engineered (by site directed mutagenesis, gene shuffling, directed evolution as described in, for example, U.S. Patent Nos. 5,837,458 and 5,723,323 both of which are herein incorporated by reference in their entirety). The invention also includes degenerate nucleic
15 acid variants of the disclosed NHP polynucleotide sequences.

Additionally contemplated are polynucleotides encoding NHP ORFs, or their functional equivalents, encoded by polynucleotide sequences that are about 99, 95, 90, or about 85 percent similar or identical to corresponding regions of
20 the nucleotide sequences of the Sequence Listing (as measured by BLAST sequence comparison analysis using, for example, the GCG sequence analysis package using standard default settings).

The invention also includes nucleic acid molecules,
25 preferably DNA molecules, that hybridize to, and are therefore the complements of, the described NHP gene nucleotide sequences. Such hybridization conditions may be highly stringent or less highly stringent, as described above. In instances where the nucleic acid molecules are
30 deoxyoligonucleotides ("DNA oligos"), such molecules are generally about 16 to about 100 bases long, or about 20 to about 80, or about 34 to about 45 bases long, or any variation or combination of sizes represented therein that

incorporate a contiguous region of sequence first disclosed in the Sequence Listing. Such oligonucleotides can be used in conjunction with the polymerase chain reaction (PCR) to screen libraries, isolate clones, and prepare cloning and sequencing templates, etc.

Alternatively, such NHP oligonucleotides can be used as hybridization probes for screening libraries, and assessing gene expression patterns (particularly using a micro array or high-throughput "chip" format). Additionally, a series of the described NHP oligonucleotide sequences, or the complements thereof, can be used to represent all or a portion of the described NHP sequences. An oligonucleotide or polynucleotide sequence first disclosed in at least a portion of one or more of the sequences of SEQ ID NOS: 1-13 can be used as a hybridization probe in conjunction with a solid support matrix/substrate (resins, beads, membranes, plastics, polymers, metal or metallized substrates, crystalline or polycrystalline substrates, etc.). Of particular note are spatially addressable arrays (*i.e.*, gene chips, microtiter plates, etc.) of oligonucleotides and polynucleotides, or corresponding oligopeptides and polypeptides, wherein at least one of the biopolymers present on the spatially addressable array comprises an oligonucleotide or polynucleotide sequence first disclosed in at least one of the sequences of SEQ ID NOS: 1-13, or an amino acid sequence encoded thereby. Methods for attaching biopolymers to, or synthesizing biopolymers on, solid support matrices, and conducting binding studies thereon are disclosed in, *inter alia*, U.S. Patent Nos. 5,700,637, 5,556,752, 5,744,305, 4,631,211, 5,445,934, 5,252,743, 4,713,326, 5,424,186, and 4,689,405 the disclosures of which are herein incorporated by reference in their entirety.

Addressable arrays comprising sequences first disclosed in SEQ ID NOS:1-13 can be used to identify and characterize the temporal and tissue specific expression of a gene. These addressable arrays incorporate oligonucleotide sequences of sufficient length to confer the required specificity, yet be within the limitations of the production technology. The length of these probes is within a range of between about 8 to about 2000 nucleotides. Preferably the probes consist of 60 nucleotides and more preferably 25 nucleotides from the sequences first disclosed in SEQ ID NOS:1-13.

For example, a series of the described oligonucleotide sequences, or the complements thereof, can be used in chip format to represent all or a portion of the described sequences. The oligonucleotides, typically between about 16 to about 40 (or any whole number within the stated range) nucleotides in length can partially overlap each other and/or the sequence may be represented using oligonucleotides that do not overlap. Accordingly, the described polynucleotide sequences shall typically comprise at least about two or three distinct oligonucleotide sequences of at least about 8 nucleotides in length that are each first disclosed in the described Sequence Listing. Such oligonucleotide sequences can begin at any nucleotide present within a sequence in the Sequence Listing and proceed in either a sense (5'-to-3') orientation vis-a-vis the described sequence or in an antisense orientation.

Microarray-based analysis allows the discovery of broad patterns of genetic activity, providing new understanding of gene functions and generating novel and unexpected insight into transcriptional processes and biological mechanisms. The use of addressable arrays comprising sequences first disclosed in SEQ ID NOS:1-13 provides detailed information about transcriptional changes involved in a specific pathway,

potentially leading to the identification of novel components or gene functions that manifest themselves as novel phenotypes.

Probes consisting of sequences first disclosed in SEQ ID NOS:1-13 can also be used in the identification, selection and validation of novel molecular targets for drug discovery. The use of these unique sequences permits the direct confirmation of drug targets and recognition of drug dependent changes in gene expression that are modulated through pathways distinct from the drugs intended target. These unique sequences therefore also have utility in defining and monitoring both drug action and toxicity.

As an example of utility, the sequences first disclosed in SEQ ID NOS:1-13 can be utilized in microarrays or other assay formats, to screen collections of genetic material from patients who have a particular medical condition. These investigations can also be carried out using the sequences first disclosed in SEQ ID NOS:1-13 *in silico* and by comparing previously collected genetic databases and the disclosed sequences using computer software known to those in the art.

Thus the sequences first disclosed in SEQ ID NOS:1-13 can be used to identify mutations associated with a particular disease and also as a diagnostic or prognostic assay.

Although the presently described sequences have been specifically described using nucleotide sequence, it should be appreciated that each of the sequences can uniquely be described using any of a wide variety of additional structural attributes, or combinations thereof. For example, a given sequence can be described by the net composition of the nucleotides present within a given region of the sequence in conjunction with the presence of one or more specific oligonucleotide sequence(s) first disclosed in the SEQ ID

NOS: 1-13. Alternatively, a restriction map specifying the relative positions of restriction endonuclease digestion sites, or various palindromic or other specific oligonucleotide sequences can be used to structurally
5 describe a given sequence. Such restriction maps, which are typically generated by widely available computer programs (e.g., the University of Wisconsin GCG sequence analysis package, SEQUENCHER 3.0, Gene Codes Corp., Ann Arbor, MI, etc.), can optionally be used in conjunction with one or more
10 discrete nucleotide sequence(s) present in the sequence that can be described by the relative position of the sequence relative to one or more additional sequence(s) or one or more restriction sites present in the disclosed sequence.

For oligonucleotide probes, highly stringent conditions
15 may refer, e.g., to washing in 6xSSC/0.05% sodium pyrophosphate at 37°C (for 14-base oligos), 48°C (for 17-base oligos), 55°C (for 20-base oligos), and 60°C (for 23-base oligos). These nucleic acid molecules may encode or act as NHP gene antisense molecules, useful, for example, in NHP
20 gene regulation (for and/or as antisense primers in amplification reactions of NHP gene nucleic acid sequences). With respect to NHP gene regulation, such techniques can be used to regulate biological functions. Further, such sequences may be used as part of ribozyme and/or triple helix
25 sequences that are also useful for NHP gene regulation.

Inhibitory antisense or double stranded oligonucleotides can additionally comprise at least one modified base moiety which is selected from the group including but not limited to 5-fluorouracil, 5-bromouracil,
30 5-chlorouracil, 5-iodouracil, hypoxanthine, xantine, 4-acetylcytosine, 5-(carboxyhydroxymethyl) uracil, 5-carboxymethylaminomethyl-2-thiouridine, 5-carboxymethylaminomethyluracil, dihydrouracil, beta-D-

galactosylqueosine, inosine, N6-isopentenyladenine,
1-methylguanine, 1-methylinosine, 2,2-dimethylguanine,
2-methyladenine, 2-methylguanine, 3-methylcytosine,
5-methylcytosine, N6-adenine, 7-methylguanine,
5 5-methylaminomethyluracil, 5-methoxyaminomethyl-2-thiouracil,
beta-D-mannosylqueosine, 5'-methoxycarboxymethyluracil,
5-methoxyuracil, 2-methylthio-N6-isopentenyladenine,
uracil-5-oxyacetic acid (v), wybutoxosine, pseudouracil,
queosine, 2-thiocytosine, 5-methyl-2-thiouracil,
10 2-thiouracil, 4-thiouracil, 5-methyluracil, uracil-
5-oxyacetic acid methylester, uracil-5-oxyacetic acid (v),
5-methyl-2-thiouracil, 3-(3-amino-3-N-2-carboxypropyl)
uracil, (acp3)w, and 2,6-diaminopurine.

The antisense oligonucleotide can also comprise at
15 least one modified sugar moiety selected from the group
including but not limited to arabinose, 2-fluoroarabinose,
xylulose, and hexose.

In yet another embodiment, the antisense
oligonucleotide will comprise at least one modified phosphate
20 backbone selected from the group consisting of a
phosphorothioate, a phosphorodithioate, a
phosphoramidothioate, a phosphoramidate, a phosphordiamidate,
a methylphosphonate, an alkyl phosphotriester, and a
formacetal or analog thereof.

25 In yet another embodiment, the antisense
oligonucleotide is an α -anomeric oligonucleotide. An α -
anomeric oligonucleotide forms specific double-stranded
hybrids with complementary RNA in which, contrary to the
usual β -units, the strands run parallel to each other
30 (Gautier et al., 1987, Nucl. Acids Res. 15:6625-6641). The
oligonucleotide is a 2'-O-methylribonucleotide (Inoue et al.,
1987, Nucl. Acids Res. 15:6131-6148), or a chimeric RNA-DNA
analogue (Inoue et al., 1987, FEBS Lett. 215:327-330).

Alternatively, double stranded RNA can be used to disrupt the expression and function of a targeted NHP.

Oligonucleotides of the invention can be synthesized by standard methods known in the art, *e.g.* by use of an automated DNA synthesizer (such as are commercially available from Biosearch, Applied Biosystems, etc.). As examples, phosphorothioate oligonucleotides can be synthesized by the method of Stein *et al.* (1988, *Nucl. Acids Res.* 16:3209), and methylphosphonate oligonucleotides can be prepared by use of controlled pore glass polymer supports (Sarin *et al.*, 1988, *Proc. Natl. Acad. Sci. U.S.A.* 85:7448-7451), etc.

Low stringency conditions are well known to those of skill in the art, and will vary predictably depending on the specific organisms from which the library and the labeled sequences are derived. For guidance regarding such conditions see, for example, Sambrook *et al.*, 1989, *Molecular Cloning, A Laboratory Manual* (and periodic updates thereof), Cold Springs Harbor Press, N.Y.; and Ausubel *et al.*, 1989, *Current Protocols in Molecular Biology*, Green Publishing Associates and Wiley Interscience, N.Y.

Alternatively, suitably labeled NHP nucleotide probes can be used to screen a human genomic library using appropriately stringent conditions or by PCR. The identification and characterization of human genomic clones is helpful for identifying polymorphisms (including, but not limited to, nucleotide repeats, microsatellite alleles, single nucleotide polymorphisms, or coding single nucleotide polymorphisms), determining the genomic structure of a given locus/allele, and designing diagnostic tests. For example, sequences derived from regions adjacent to the intron/exon boundaries of the human gene can be used to design primers for use in amplification assays to detect mutations within

the exons, introns, splice sites (e.g., splice acceptor and/or donor sites), etc.; that can be used in diagnostics and pharmacogenomics.

Further, a NHP gene homolog can be isolated from
5 nucleic acid from an organism of interest by performing PCR using two degenerate or "wobble" oligonucleotide primer pools designed on the basis of amino acid sequences within the NHP products disclosed herein. The template for the reaction may be total RNA, mRNA, and/or cDNA obtained by reverse
10 transcription of mRNA prepared from human or non-human cell lines or tissue known or suspected to express an allele of a NHP gene. The PCR product can be subcloned and sequenced to ensure that the amplified sequences represent the sequence of the desired NHP gene. The PCR fragment can then be used to
15 isolate a full length cDNA clone by a variety of methods. For example, the amplified fragment can be labeled and used to screen a cDNA library, such as a bacteriophage cDNA library. Alternatively, the labeled fragment can be used to isolate genomic clones via the screening of a genomic
20 library.

PCR technology can also be used to isolate full length cDNA sequences. For example, RNA can be isolated, following standard procedures, from an appropriate cellular or tissue source (i.e., one known, or suspected, to express a NHP
25 gene). A reverse transcription (RT) reaction can be performed on the RNA using an oligonucleotide primer specific for the most 5' end of the amplified fragment for the priming of first strand synthesis. The resulting RNA/DNA hybrid may then be "tailed" using a standard terminal transferase
30 reaction, the hybrid may be digested with RNase H, and second strand synthesis may then be primed with a complementary primer. Thus, cDNA sequences upstream of the amplified

fragment can be isolated. For a review of cloning strategies that can be used, see *e.g.*, Sambrook et al., 1989, *supra*.

A cDNA encoding a mutant NHP gene can be isolated, for example, by using PCR. In this case, the first cDNA strand
5 may be synthesized by hybridizing an oligo-dT oligonucleotide to mRNA isolated from tissue known or suspected to be expressed in an individual putatively carrying a mutant NHP allele, and by extending the new strand with reverse transcriptase. The second strand of the cDNA is then
10 synthesized using an oligonucleotide that hybridizes specifically to the 5' end of the normal gene. Using these two primers, the product is then amplified via PCR, optionally cloned into a suitable vector, and subjected to DNA sequence analysis through methods well known to those of
15 skill in the art. By comparing the DNA sequence of the mutant NHP allele to that of a corresponding normal NHP allele, the mutation(s) responsible for the loss or alteration of function of the mutant NHP gene product can be ascertained.

20 Alternatively, a genomic library can be constructed using DNA obtained from an individual suspected of or known to carry a mutant NHP allele (*e.g.*, a person manifesting a NHP-associated phenotype such as, for example, obesity, high blood pressure, connective tissue disorders, infertility,
25 etc.), or a cDNA library can be constructed using RNA from a tissue known, or suspected, to express a mutant NHP allele. A normal NHP gene, or any suitable fragment thereof, can then be labeled and used as a probe to identify the corresponding mutant NHP allele in such libraries. Clones containing
30 mutant NHP gene sequences can then be purified and subjected to sequence analysis according to methods well known to those skilled in the art.

Additionally, an expression library can be constructed utilizing cDNA synthesized from, for example, RNA isolated from a tissue known, or suspected, to express a mutant NHP allele in an individual suspected of or known to carry such a mutant allele. In this manner, gene products made by the putatively mutant tissue can be expressed and screened using standard antibody screening techniques in conjunction with antibodies raised against a normal NHP product, as described below. (For screening techniques, see, for example, Harlow, E. and Lane, eds., 1988, "Antibodies: A Laboratory Manual", Cold Spring Harbor Press, Cold Spring Harbor, NY.)

Additionally, screening can be accomplished by screening with labeled NHP fusion proteins, such as, for example, alkaline phosphatase-NHP or NHP-alkaline phosphatase fusion proteins. In cases where a NHP mutation results in an expressed gene product with altered function (e.g., as a result of a missense or a frameshift mutation), polyclonal antibodies to a NHP are likely to cross-react with a corresponding mutant NHP gene product. Library clones detected via their reaction with such labeled antibodies can be purified and subjected to sequence analysis according to methods well known in the art.

The invention also encompasses (a) DNA vectors that contain any of the foregoing NHP coding sequences and/or their complements (i.e., antisense); (b) DNA expression vectors that contain any of the foregoing NHP coding sequences operatively associated with a regulatory element that directs the expression of the coding sequences (for example, baculo virus as described in U.S. Patent No. 5,869,336 herein incorporated by reference); (c) genetically engineered host cells that contain any of the foregoing NHP coding sequences operatively associated with a regulatory element that directs the expression of the coding sequences

in the host cell; and (d) genetically engineered host cells that express an endogenous NHP gene under the control of an exogenously introduced regulatory element (i.e., gene activation). As used herein, regulatory elements include, but are not limited to, inducible and non-inducible promoters, enhancers, operators and other elements known to those skilled in the art that drive and regulate expression. Such regulatory elements include but are not limited to the cytomegalovirus (hCMV) immediate early gene, regulatable, viral elements (particularly retroviral LTR promoters), the early or late promoters of SV40 adenovirus, the *lac* system, the *trp* system, the *TAC* system, the *TRC* system, the major operator and promoter regions of phage lambda, the control regions of fd coat protein, the promoter for 3-phosphoglycerate kinase (PGK), the promoters of acid phosphatase, and the promoters of the yeast α -mating factors.

The present invention also encompasses antibodies and anti-idiotypic antibodies (including Fab fragments), antagonists and agonists of the NHP, as well as compounds or nucleotide constructs that inhibit expression of a NHP gene (transcription factor inhibitors, antisense and ribozyme molecules, or gene or regulatory sequence replacement constructs), or promote the expression of a NHP (e.g., expression constructs in which NHP coding sequences are operatively associated with expression control elements such as promoters, promoter/enhancers, etc.).

The NHPs or NHP peptides, NHP fusion proteins, NHP nucleotide sequences, antibodies, antagonists and agonists can be useful for the detection of mutant NHPs or inappropriately expressed NHPs for the diagnosis of disease. The NHP proteins or peptides, NHP fusion proteins, NHP nucleotide sequences, host cell expression systems, antibodies, antagonists, agonists and genetically engineered

cells and animals can be used for screening for drugs (or high throughput screening of combinatorial libraries) effective in the treatment of the symptomatic or phenotypic manifestations of perturbing the normal function of NHP in the body. The use of engineered host cells and/or animals may offer an advantage in that such systems allow not only for the identification of compounds that bind to the endogenous receptor for an NHP, but can also identify compounds that trigger NHP-mediated activities or pathways.

Finally, the NHP products can be used as therapeutics. For example, soluble derivatives such as NHP peptides/domains corresponding to NHPs, NHP fusion protein products (especially NHP-Ig fusion proteins, *i.e.*, fusions of a NHP, or a domain of a NHP, to an IgFc), NHP antibodies and anti-idiotypic antibodies (including Fab fragments), antagonists or agonists (including compounds that modulate or act on downstream targets in a NHP-mediated pathway) can be used to directly treat diseases or disorders. For instance, the administration of an effective amount of soluble NHP, or a NHP-IgFc fusion protein or an anti-idiotypic antibody (or its Fab) that mimics the NHP could activate or effectively antagonize the endogenous NHP receptor. Nucleotide constructs encoding such NHP products can be used to genetically engineer host cells to express such products *in vivo*; these genetically engineered cells function as "bioreactors" in the body delivering a continuous supply of a NHP, a NHP peptide, or a NHP fusion protein to the body. Nucleotide constructs encoding functional NHPs, mutant NHPs, as well as antisense and ribozyme molecules can also be used in "gene therapy" approaches for the modulation of NHP expression. Thus, the invention also encompasses pharmaceutical formulations and methods for treating biological disorders.

Various aspects of the invention are described in greater detail in the subsections below.

5.1 THE NHP SEQUENCES

5 The cDNA sequences and the corresponding deduced amino acid sequences of the described NHPs are presented in the Sequence Listing. The NHP nucleotides were obtained from clustered human ESTs, human thyroid RACE products, and cDNAs from kidney and thyroid libraries (Edge Biosystems, 10 Gaithersburg, MD). The described NHPs are similar to eucaryotic sulfate transporters and human pendrin. A particularly similar sulfate transporter has been associated with diastrophic dysplasia which has been associated with dwarfism, inherited chondrodysplasia, and osseous dysplasia 15 I. Accordingly, the described NHPs can be useful in detecting and treating such conditions.

A polymorphism was detected consisting of an A-T transversion at the sequence region represented by, for example, nucleotide number 1454 of SEQ ID NO:1 which can 20 result in a corresponding E or V being present at amino acid position 485 of, for example, SEQ ID NO:2.

Transporters and transporter related multidrug resistance (MDR) sequences, as well as uses and applications that are germane to the described NHPs, are described in U.S. 25 Patents Nos. 5,198,344 and 5,866,699 which are herein incorporated by reference in their entirety.

An additional application of the described novel human polynucleotide sequences is their use in the molecular mutagenesis/evolution of proteins that are at least partially 30 encoded by the described novel sequences using, for example, polynucleotide shuffling or related methodologies. Such approaches are described in U.S. Patents Nos. 5,830,721 and

5,837,458 which are herein incorporated by reference in their entirety.

NHP gene products can also be expressed in transgenic animals. Animals of any species, including, but not limited to, worms, mice, rats, rabbits, guinea pigs, pigs, micro-pigs, birds, goats, and non-human primates, e.g., baboons, monkeys, and chimpanzees may be used to generate NHP transgenic animals.

Any technique known in the art may be used to introduce a NHP transgene into animals to produce the founder lines of transgenic animals. Such techniques include, but are not limited to pronuclear microinjection (Hoppe, P.C. and Wagner, T.E., 1989, U.S. Pat. No. 4,873,191); retrovirus mediated gene transfer into germ lines (Van der Putten *et al.*, 1985, Proc. Natl. Acad. Sci., USA 82:6148-6152); gene targeting in embryonic stem cells (Thompson *et al.*, 1989, Cell 56:313-321); electroporation of embryos (Lo, 1983, Mol Cell. Biol. 3:1803-1814); and sperm-mediated gene transfer (Lavitrano *et al.*, 1989, Cell 57:717-723); etc. For a review of such techniques, see Gordon, 1989, Transgenic Animals, Intl. Rev. Cytol. 115:171-229, which is incorporated by reference herein in its entirety.

The present invention provides for transgenic animals that carry the NHP transgene in all their cells, as well as animals which carry the transgene in some, but not all their cells, i.e., mosaic animals or somatic cell transgenic animals. The transgene may be integrated as a single transgene or in concatamers, e.g., head-to-head tandems or head-to-tail tandems. The transgene may also be selectively introduced into and activated in a particular cell type by following, for example, the teaching of Lasko *et al.*, 1992, Proc. Natl. Acad. Sci. USA 89:6232-6236. The regulatory

sequences required for such a cell-type specific activation will depend upon the particular cell type of interest, and will be apparent to those of skill in the art.

When it is desired that a NHP transgene be integrated
5 into the chromosomal site of the endogenous NHP gene, gene targeting is preferred. Briefly, when such a technique is to be utilized, vectors containing some nucleotide sequences homologous to the endogenous NHP gene are designed for the purpose of integrating, via homologous recombination with
10 chromosomal sequences, into and disrupting the function of the nucleotide sequence of the endogenous NHP gene (*i.e.*, "knockout" animals).

The transgene can also be selectively introduced into a particular cell type, thus inactivating the endogenous NHP
15 gene in only that cell type, by following, for example, the teaching of Gu *et al.*, 1994, *Science*, 265:103-106. The regulatory sequences required for such a cell-type specific inactivation will depend upon the particular cell type of interest, and will be apparent to those of skill in the art.

20 Once transgenic animals have been generated, the expression of the recombinant NHP gene may be assayed utilizing standard techniques. Initial screening may be accomplished by Southern blot analysis or PCR techniques to analyze animal tissues to assay whether integration of the
25 transgene has taken place. The level of mRNA expression of the transgene in the tissues of the transgenic animals may also be assessed using techniques which include but are not limited to Northern blot analysis of tissue samples obtained from the animal, *in situ* hybridization analysis, and RT-PCR.
30 Samples of NHP gene-expressing tissue, may also be evaluated immunocytochemically using antibodies specific for the NHP transgene product.

5.2 NHPS AND NHP POLYPEPTIDES

NHPS, polypeptides, peptide fragments, mutated, truncated, or deleted forms of the NHPS, and/or NHP fusion proteins can be prepared for a variety of uses. These uses include but are not limited to the generation of antibodies, as reagents in diagnostic assays, the identification of other cellular gene products related to a NHP, as reagents in assays for screening for compounds that can be used as pharmaceutical reagents useful in the therapeutic treatment of mental, biological, or medical disorders and diseases. Given the similarity information and expression data, the described NHPS can be targeted (by drugs, oligos, antibodies, etc,) in order to treat disease, or to therapeutically augment the efficacy of, for example, chemotherapeutic agents used in the treatment of breast or prostate cancer.

The Sequence Listing discloses the amino acid sequences encoded by the described NHP genes. The NHPS typically display have initiator methionines in DNA sequence contexts consistent with a translation initiation site.

The NHP amino acid sequences of the invention include the amino acid sequence presented in the Sequence Listing as well as analogues and derivatives thereof. Further, corresponding NHP homologues from other species are encompassed by the invention. In fact, any NHP protein encoded by the NHP nucleotide sequences described above are within the scope of the invention, as are any novel polynucleotide sequences encoding all or any novel portion of an amino acid sequence presented in the Sequence Listing. The degenerate nature of the genetic code is well known, and, accordingly, each amino acid presented in the Sequence Listing, is generically representative of the well known nucleic acid "triplet" codon, or in many cases codons, that can encode the amino acid. As such, as contemplated herein,

the amino acid sequences presented in the Sequence Listing, when taken together with the genetic code (see, for example, Table 4-1 at page 109 of "Molecular Cell Biology", 1986, J. Darnell et al. eds., Scientific American Books, New York, NY, herein incorporated by reference) are generically representative of all the various permutations and combinations of nucleic acid sequences that can encode such amino acid sequences.

The invention also encompasses proteins that are functionally equivalent to the NHPs encoded by the presently described nucleotide sequences as judged by any of a number of criteria, including, but not limited to, the ability to bind and cleave a substrate of a NHP, or the ability to effect an identical or complementary downstream pathway, or a change in cellular metabolism (e.g., proteolytic activity, ion flux, tyrosine phosphorylation, etc.). Such functionally equivalent NHP proteins include, but are not limited to, additions or substitutions of amino acid residues within the amino acid sequence encoded by the NHP nucleotide sequences described above, but which result in a silent change, thus producing a functionally equivalent gene product. Amino acid substitutions may be made on the basis of similarity in polarity, charge, solubility, hydrophobicity, hydrophilicity, and/or the amphipathic nature of the residues involved. For example, nonpolar (hydrophobic) amino acids include alanine, leucine, isoleucine, valine, proline, phenylalanine, tryptophan, and methionine; polar neutral amino acids include glycine, serine, threonine, cysteine, tyrosine, asparagine, and glutamine; positively charged (basic) amino acids include arginine, lysine, and histidine; and negatively charged (acidic) amino acids include aspartic acid and glutamic acid.

A variety of host-expression vector systems can be used to express the NHP nucleotide sequences of the invention.

Where, as in the present instance, the NHP peptide or polypeptide is thought to be membrane protein, the hydrophobic regions of the protein can be excised and the resulting soluble peptide or polypeptide can be recovered from the culture media. Such expression systems also encompass engineered host cells that express a NHP, or functional equivalent, *in situ*. Purification or enrichment of a NHP from such expression systems can be accomplished using appropriate detergents and lipid micelles and methods well known to those skilled in the art. However, such engineered host cells themselves may be used in situations where it is important not only to retain the structural and functional characteristics of the NHP, but to assess biological activity, *e.g.*, in drug screening assays.

The expression systems that may be used for purposes of the invention include but are not limited to microorganisms such as bacteria (*e.g.*, *E. coli*, *B. subtilis*) transformed with recombinant bacteriophage DNA, plasmid DNA or cosmid DNA expression vectors containing NHP nucleotide sequences; yeast (*e.g.*, *Saccharomyces*, *Pichia*) transformed with recombinant yeast expression vectors containing NHP nucleotide sequences; insect cell systems infected with recombinant virus expression vectors (*e.g.*, baculovirus) containing NHP sequences; plant cell systems infected with recombinant virus expression vectors (*e.g.*, cauliflower mosaic virus, CaMV; tobacco mosaic virus, TMV) or transformed with recombinant plasmid expression vectors (*e.g.*, Ti plasmid) containing NHP nucleotide sequences; or mammalian cell systems (*e.g.*, COS, CHO, BHK, 293, 3T3) harboring recombinant expression constructs containing promoters derived from the genome of mammalian cells (*e.g.*, metallothionein promoter) or from mammalian

viruses (e.g., the adenovirus late promoter; the vaccinia virus 7.5K promoter).

In bacterial systems, a number of expression vectors may be advantageously selected depending upon the use intended for the NHP product being expressed. For example, when a large quantity of such a protein is to be produced for the generation of pharmaceutical compositions of or containing NHP, or for raising antibodies to a NHP, vectors that direct the expression of high levels of fusion protein products that are readily purified may be desirable. Such vectors include, but are not limited, to the *E. coli* expression vector pUR278 (Ruther et al., 1983, EMBO J. 2:1791), in which a NHP coding sequence may be ligated individually into the vector in frame with the *lacZ* coding region so that a fusion protein is produced; pIN vectors (Inouye & Inouye, 1985, Nucleic Acids Res. 13:3101-3109; Van Heeke & Schuster, 1989, J. Biol. Chem. 264:5503-5509); and the like. pGEX vectors (Pharmacia or American Type Culture Collection) can also be used to express foreign polypeptides as fusion proteins with glutathione S-transferase (GST). In general, such fusion proteins are soluble and can easily be purified from lysed cells by adsorption to glutathione-agarose beads followed by elution in the presence of free glutathione. The pGEX vectors are designed to include thrombin or factor Xa protease cleavage sites so that the cloned target gene product can be released from the GST moiety.

In an insect system, *Autographa californica* nuclear polyhidrosis virus (AcNPV) is used as a vector to express foreign genes. The virus grows in *Spodoptera frugiperda* cells. A NHP gene coding sequence may be cloned individually into non-essential regions (for example the polyhedrin gene)

of the virus and placed under control of an AcNPV promoter (for example the polyhedrin promoter). Successful insertion of NHP gene coding sequence will result in inactivation of the polyhedrin gene and production of non-occluded

5 recombinant virus (i.e., virus lacking the proteinaceous coat coded for by the polyhedrin gene). These recombinant viruses are then used to infect *Spodoptera frugiperda* cells in which the inserted gene is expressed (e.g., see Smith et al., 1983, J. Virol. 46:584; Smith, U.S. Patent No. 4,215,051).

10 In mammalian host cells, a number of viral-based expression systems may be utilized. In cases where an adenovirus is used as an expression vector, the NHP nucleotide sequence of interest may be ligated to an adenovirus transcription/translation control complex, e.g.,
15 the late promoter and tripartite leader sequence. This chimeric gene may then be inserted in the adenovirus genome by *in vitro* or *in vivo* recombination. Insertion in a non-essential region of the viral genome (e.g., region E1 or E3) will result in a recombinant virus that is viable and capable
20 of expressing a NHP product in infected hosts (e.g., See Logan & Shenk, 1984, Proc. Natl. Acad. Sci. USA 81:3655-3659). Specific initiation signals may also be required for efficient translation of inserted NHP nucleotide sequences. These signals include the ATG initiation codon and adjacent
25 sequences. In cases where an entire NHP gene or cDNA, including its own initiation codon and adjacent sequences, is inserted into the appropriate expression vector, no additional translational control signals may be needed. However, in cases where only a portion of a NHP coding
30 sequence is inserted, exogenous translational control signals, including, perhaps, the ATG initiation codon, must be provided. Furthermore, the initiation codon must be in

phase with the reading frame of the desired coding sequence to ensure translation of the entire insert. These exogenous translational control signals and initiation codons can be of a variety of origins, both natural and synthetic. The efficiency of expression may be enhanced by the inclusion of appropriate transcription enhancer elements, transcription terminators, etc. (See Bitter *et al.*, 1987, *Methods in Enzymol.* 153:516-544).

In addition, a host cell strain may be chosen that modulates the expression of the inserted sequences, or modifies and processes the gene product in the specific fashion desired. Such modifications (e.g., glycosylation) and processing (e.g., cleavage) of protein products may be important for the function of the protein. Different host cells have characteristic and specific mechanisms for the post-translational processing and modification of proteins and gene products. Appropriate cell lines or host systems can be chosen to ensure the correct modification and processing of the foreign protein expressed. To this end, eukaryotic host cells which possess the cellular machinery for proper processing of the primary transcript, glycosylation, and phosphorylation of the gene product may be used. Such mammalian host cells include, but are not limited to, CHO, VERO, BHK, HeLa, COS, MDCK, 293, 3T3, WI38, and in particular, human cell lines.

For long-term, high-yield production of recombinant proteins, stable expression is preferred. For example, cell lines which stably express the NHP sequences described above can be engineered. Rather than using expression vectors which contain viral origins of replication, host cells can be transformed with DNA controlled by appropriate expression control elements (e.g., promoter, enhancer sequences, transcription terminators, polyadenylation sites, etc.), and

a selectable marker. Following the introduction of the foreign DNA, engineered cells may be allowed to grow for 1-2 days in an enriched media, and then are switched to a selective media. The selectable marker in the recombinant
5 plasmid confers resistance to the selection and allows cells to stably integrate the plasmid into their chromosomes and grow to form foci which in turn can be cloned and expanded into cell lines. This method may advantageously be used to engineer cell lines which express the NHP product. Such
10 engineered cell lines may be particularly useful in screening and evaluation of compounds that affect the endogenous activity of the NHP product.

A number of selection systems may be used, including but not limited to the herpes simplex virus thymidine kinase
15 (Wigler, *et al.*, 1977, *Cell* 11:223), hypoxanthine-guanine phosphoribosyltransferase (Szybalska & Szybalski, 1962, *Proc. Natl. Acad. Sci. USA* 48:2026), and adenine phosphoribosyltransferase (Lowy, *et al.*, 1980, *Cell* 22:817) genes can be employed in *tk*⁻, *hgp*⁻ or *aprt*⁻ cells,
20 respectively. Also, antimetabolite resistance can be used as the basis of selection for the following genes: *dhfr*, which confers resistance to methotrexate (Wigler, *et al.*, 1980, *Natl. Acad. Sci. USA* 77:3567; O'Hare, *et al.*, 1981, *Proc. Natl. Acad. Sci. USA* 78:1527); *gpt*, which confers resistance
25 to mycophenolic acid (Mulligan & Berg, 1981, *Proc. Natl. Acad. Sci. USA* 78:2072); *neo*, which confers resistance to the aminoglycoside G-418 (Colberre-Garapin, *et al.*, 1981, *J. Mol. Biol.* 150:1); and *hygro*, which confers resistance to hygromycin (Santerre, *et al.*, 1984, *Gene* 30:147).

30 Alternatively, any fusion protein can be readily purified by utilizing an antibody specific for the fusion protein being expressed. For example, a system described by

Janknecht et al. allows for the ready purification of non-denatured fusion proteins expressed in human cell lines (Janknecht, et al., 1991, Proc. Natl. Acad. Sci. USA 88:8972-8976). In this system, the gene of interest is subcloned
5 into a vaccinia recombination plasmid such that the gene's open reading frame is translationally fused to an amino-terminal tag consisting of six histidine residues. Extracts from cells infected with recombinant vaccinia virus are loaded onto Ni²⁺-nitriloacetic acid-agarose columns and
10 histidine-tagged proteins are selectively eluted with imidazole-containing buffers.

Also encompassed by the present invention are fusion proteins that direct the NHP to a target organ and/or facilitate transport across the membrane into the cytosol.
15 Conjugation of NHPs to antibody molecules or their Fab fragments could be used to target cells bearing a particular epitope. Attaching the appropriate signal sequence to the NHP would also transport the NHP to the desired location within the cell. Alternatively targeting of NHP or its
20 nucleic acid sequence might be achieved using liposome or lipid complex based delivery systems. Such technologies are described in "Liposomes: A Practical Approach", New, RRC ed., Oxford University Press, New York and in U.S. Patents Nos. 4,594,595, 5,459,127, 5,948,767 and 6,110,490 and their
25 respective disclosures which are herein incorporated by reference in their entirety. Additionally embodied are novel protein constructs engineered in such a way that they facilitate transport of the NHP to the target site or desired organ. This goal may be achieved by coupling of the NHP to a
30 cytokine or other ligand that provides targeting specificity, and/or to a protein transducing domain (see generally U.S. applications Ser. No. 60/111,701 and 60/056,713, both of which are herein incorporated by reference, for examples of

such transducing sequences) to facilitate passage across cellular membranes if needed and can optionally be engineered to include nuclear localization sequences when desired.

5 5.3 ANTIBODIES TO NHP PRODUCTS

Antibodies that specifically recognize one or more epitopes of a NHP, or epitopes of conserved variants of a NHP, or peptide fragments of a NHP are also encompassed by the invention. Such antibodies include but are not limited
10 to polyclonal antibodies, monoclonal antibodies (mAbs), humanized or chimeric antibodies, single chain antibodies, Fab fragments, F(ab')₂ fragments, fragments produced by a Fab expression library, anti-idiotypic (anti-Id) antibodies, and epitope-binding fragments of any of the above.

15 The antibodies of the invention may be used, for example, in the detection of NHP in a biological sample and may, therefore, be utilized as part of a diagnostic or prognostic technique whereby patients may be tested for abnormal amounts of NHP. Such antibodies may also be
20 utilized in conjunction with, for example, compound screening schemes for the evaluation of the effect of test compounds on expression and/or activity of a NHP gene product. Additionally, such antibodies can be used in conjunction gene therapy to, for example, evaluate the normal and/or
25 engineered NHP-expressing cells prior to their introduction into the patient. Such antibodies may additionally be used as a method for the inhibition of abnormal NHP activity. Thus, such antibodies may, therefore, be utilized as part of treatment methods.

30 For the production of antibodies, various host animals may be immunized by injection with a NHP, an NHP peptide (e.g., one corresponding to a functional domain of an NHP), truncated NHP polypeptides (NHP in which one or more domains

have been deleted), functional equivalents of the NHP or mutated variant of the NHP. Such host animals may include but are not limited to pigs, rabbits, mice, goats, and rats, to name but a few. Various adjuvants may be used to increase the immunological response, depending on the host species, including but not limited to Freund's adjuvant (complete and incomplete), mineral salts such as aluminum hydroxide or aluminum phosphate, surface active substances such as lysolecithin, pluronic polyols, polyanions, peptides, oil emulsions, and potentially useful human adjuvants such as BCG (bacille Calmette-Guerin) and *Corynebacterium parvum*. Alternatively, the immune response could be enhanced by combination and or coupling with molecules such as keyhole limpet hemocyanin, tetanus toxoid, diphtheria toxoid, ovalbumin, cholera toxin or fragments thereof. Polyclonal antibodies are heterogeneous populations of antibody molecules derived from the sera of the immunized animals.

Monoclonal antibodies, which are homogeneous populations of antibodies to a particular antigen, can be obtained by any technique which provides for the production of antibody molecules by continuous cell lines in culture. These include, but are not limited to, the hybridoma technique of Kohler and Milstein, (1975, Nature 256:495-497; and U.S. Patent No. 4,376,110); the human B-cell hybridoma technique (Kosbor et al., 1983, Immunology Today 4:72; Cole et al., 1983, Proc. Natl. Acad. Sci. USA 80:2026-2030), and the EBV-hybridoma technique (Cole et al., 1985, Monoclonal Antibodies And Cancer Therapy, Alan R. Liss, Inc., pp. 77-96). Such antibodies may be of any immunoglobulin class including IgG, IgM, IgE, IgA, IgD and any subclass thereof. The hybridoma producing the mAb of this invention may be cultivated *in vitro* or *in vivo*. Production of high titers of

mAbs *in vivo* makes this the presently preferred method of production.

In addition, techniques developed for the production of "chimeric antibodies" (Morrison *et al.*, 1984, Proc. Natl. Acad. Sci., 81:6851-6855; Neuberger *et al.*, 1984, Nature, 312:604-608; Takeda *et al.*, 1985, Nature, 314:452-454) by splicing the genes from a mouse antibody molecule of appropriate antigen specificity together with genes from a human antibody molecule of appropriate biological activity can be used. A chimeric antibody is a molecule in which different portions are derived from different animal species, such as those having a variable region derived from a murine mAb and a human immunoglobulin constant region. Such technologies are described in U.S. Patents Nos. 6,075,181 and 5,877,397 and their respective disclosures which are herein incorporated by reference in their entirety. Also encompassed by the present invention is the use of fully humanized monoclonal antibodies as described in US Patent No. 6,150,584 and respective disclosures which are herein incorporated by reference in their entirety.

Alternatively, techniques described for the production of single chain antibodies (U.S. Patent 4,946,778; Bird, 1988, Science 242:423-426; Huston *et al.*, 1988, Proc. Natl. Acad. Sci. USA 85:5879-5883; and Ward *et al.*, 1989, Nature 341:544-546) can be adapted to produce single chain antibodies against NHP gene products. Single chain antibodies are formed by linking the heavy and light chain fragments of the Fv region via an amino acid bridge, resulting in a single chain polypeptide.

Antibody fragments which recognize specific epitopes may be generated by known techniques. For example, such fragments include, but are not limited to: the F(ab')₂

fragments which can be produced by pepsin digestion of the antibody molecule and the Fab fragments which can be generated by reducing the disulfide bridges of the F(ab')₂ fragments. Alternatively, Fab expression libraries may be constructed (Huse et al., 1989, Science, 246:1275-1281) to allow rapid and easy identification of monoclonal Fab fragments with the desired specificity.

Antibodies to a NHP can, in turn, be utilized to generate anti-idiotypic antibodies that "mimic" a given NHP, using techniques well known to those skilled in the art. (See, e.g., Greenspan & Bona, 1993, FASEB J 7(5):437-444; and Nissinoff, 1991, J. Immunol. 147(8):2429-2438). For example antibodies which bind to a NHP domain and competitively inhibit the binding of NHP to its cognate receptor can be used to generate anti-idiotypes that "mimic" the NHP and, therefore, bind and activate or neutralize a receptor. Such anti-idiotypic antibodies or Fab fragments of such anti-idiotypes can be used in therapeutic regimens involving a NHP mediated pathway.

The present invention is not to be limited in scope by the specific embodiments described herein, which are intended as single illustrations of individual aspects of the invention, and functionally equivalent methods and components are within the scope of the invention. Indeed, various modifications of the invention, in addition to those shown and described herein will become apparent to those skilled in the art from the foregoing description. Such modifications are intended to fall within the scope of the appended claims. All cited publications, patents, and patent applications are herein incorporated by reference in their entirety.

WHAT IS CLAIMED IS:

1. An isolated nucleic acid molecule comprising
at a nucleotide sequence encoding an amino acid sequence
5 drawn from the group consisting of SEQ ID NOS: 2, 4, 6, 8,
10, and 12.
2. An isolated nucleic acid molecule comprising
a nucleotide sequence that:
10 (a) encodes the amino acid sequence shown in SEQ
ID NO: 10; and
(b) hybridizes under stringent conditions to the
nucleotide sequence of SEQ ID NO:9 or the
complement thereof.
15
3. An isolated nucleic acid molecule comprising
a nucleotide sequence that encodes the amino acid sequence
shown in SEQ ID NO:10.
- 20 4. An isolated nucleic acid molecule comprising
a nucleotide sequence that encodes the amino acid sequence
shown in SEQ ID NO:4.
- 25 5. An isolated nucleic acid molecule comprising
a nucleotide sequence that encodes the amino acid sequence
shown in SEQ ID NO:2.

SEQUENCE LISTING

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 405 410 415
 Leu Lys Gly Met Leu Ile Gln Phe Arg Asp Leu Lys Lys Tyr Trp Asn
 420 425 430
 Val Asp Lys Ile Asp Trp Gly Ile Trp Val Ser Thr Tyr Val Phe Thr
 435 440 445
 Ile Cys Phe, Ala Ala Asn Val Gly Leu Leu Phe Gly Val Val Cys Thr
 450 455 460
 Ile Ala Ile Val Ile Gly Arg Phe Pro Arg Ala Met Thr Val Ser Ile
 465 470 475 480
 Lys Asn Met Lys Glu Met Glu Phe Lys Val Lys Thr Glu Met Asp Ser
 485 490 495
 Glu Thr Leu Gln Gln Val Lys Ile Ile Ser Ile Asn Asn Pro Leu Val
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 Phe Leu Asn Ala Lys Lys Phe Tyr Thr Asp Leu Met Asn Met Ile Gln
 515 520 525
 Lys Glu Asn Ala Cys Asn Gln Pro Leu Asp Asp Ile Ser Lys Cys Glu
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 Gln Asn Thr Leu Leu Asn Ser Leu Ser Asn Gly Asn Cys Asn Glu Glu
 545 550 555 560
 Ala Ser Gln Ser Cys Pro Asn Glu Lys Cys Tyr Leu Ile Leu Asp Cys
 565 570 575
 Ser Gly Phe Thr Phe Phe Asp Tyr Ser Gly Val Ser Met Leu Val Glu
 580 585 590
 Val Tyr Met Asp Cys Lys Gly Arg Ser Val Asp Val Leu Leu Ala His
 595 600 605
 Cys Thr Ala Ser Leu Ile Lys Ala Met Thr Tyr Tyr Gly Asn Leu Asp
 610 615 620
 Ser Glu Lys Pro Ile Phe Phe Glu Ser Val Ser Ala Ala Ile Ser His
 625 630 635 640
 Ile His Ser Asn Lys Met Glu Ser Arg Ser Val Ser His Ala Gly Val
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<211> 1866

<212> DNA

<213> homo sapiens

<400> 3

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gaaatgcaaa ggatccacgt tgctgcagca gtttccttct tgggaggtgt gattcaggtg	480
gccatgtttg tgctgcaact gggcagtgcc acatttgttg tcacagagcc tgtgatcagc	540
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atgaaaatgc catatatatc cggaccactt ggattctttt atatttatgc atatgttttt	660

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<210> 4

<211> 621

<212> PRT

<213> homo sapiens

<400> 4

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  20          25          30
Pro Ile Leu Asp Trp Ala Pro His Tyr Asn Leu Lys Glu Asn Leu Leu
  35          40          45
Pro Asp Thr Val Ser Gly Ile Met Leu Ala Val Gln Gln Val Thr Gln
  50          55          60
Gly Leu Ala Phe Ala Val Leu Ser Ser Val His Pro Val Phe Gly Leu
  65          70          75          80
Tyr Gly Ser Leu Phe Pro Ala Ile Ile Tyr Ala Ile Phe Gly Met Gly
  85          90          95
His His Val Ala Thr Gly Thr Phe Ala Leu Thr Ser Leu Ile Ser Ala
 100          105          110
Asn Ala Val Glu Arg Ile Val Pro Gln Asn Met Gln Asn Leu Thr Thr
 115          120          125
Gln Ser Asn Thr Ser Val Leu Gly Leu Ser Asp Phe Glu Met Gln Arg
 130          135          140
Ile His Val Ala Ala Val Ser Phe Leu Gly Gly Val Ile Gln Val
 145          150          155          160
Ala Met Phe Val Leu Gln Leu Gly Ser Ala Thr Phe Val Val Thr Glu
 165          170          175
Pro Val Ile Ser Ala Met Thr Thr Gly Ala Ala Thr His Val Val Thr
 180          185          190
Ser Gln Val Lys Tyr Leu Leu Gly Met Lys Met Pro Tyr Ile Ser Gly
 195          200          205
Pro Leu Gly Phe Phe Tyr Ile Tyr Ala Tyr Val Phe Glu Asn Ile Lys
 210          215          220

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Ser Val Arg Leu Glu Ala Leu Leu Leu Ser Leu Leu Ser Ile Val Val
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Leu Val Leu Val Lys Glu Leu Asn Glu Gln Phe Lys Arg Lys Ile Lys
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Val Val Leu Pro Val Asp Leu Val Leu Ile Ile Ala Ala Ser Phe Ala
                260                265                270
Cys Tyr Cys Thr Asn Met Glu Asn Thr Tyr Gly Leu Glu Val Val Gly
275                280                285
His Ile Pro Gln Gly Ile Pro Ser Pro Arg Ala Pro Pro Met Asn Ile
290                295                300
Leu Ser Ala Val Ile Thr Glu Ala Phe Gly Val Ala Leu Val Gly Tyr
305                310                315                320
Val Ala Ser Leu Ala Leu Ala Gln Gly Ser Ala Lys Lys Phe Lys Tyr
                325                330                335
Ser Ile Asp Asp Asn Gln Glu Phe Leu Ala His Gly Leu Ser Asn Ile
                340                345                350
Val Ser Ser Phe Phe Phe Cys Ile Pro Ser Ala Ala Ala Met Gly Arg
                355                360                365
Thr Ala Gly Leu Tyr Ser Thr Gly Ala Lys Thr Gln Val Ala Cys Leu
370                375                380
Ile Ser Cys Ile Phe Val Leu Ile Val Ile Tyr Ala Ile Gly Pro Leu
385                390                395                400
Leu Tyr Trp Leu Pro Met Cys Val Leu Ala Ser Ile Ile Val Val Gly
                405                410                415
Leu Lys Gly Met Leu Ile Gln Phe Arg Asp Leu Lys Lys Tyr Trp Asn
                420                425                430
Val Asp Lys Ile Asp Trp Gly Thr Leu Gln Gln Val Lys Ile Ile Ser
                435                440                445
Ile Asn Asn Pro Leu Val Phe Leu Asn Ala Lys Lys Phe Tyr Thr Asp
450                455                460
Leu Met Asn Met Ile Gln Lys Glu Asn Ala Cys Asn Gln Pro Leu Asp
465                470                475                480
Asp Ile Ser Lys Cys Glu Gln Asn Thr Leu Leu Asn Ser Leu Ser Asn
                485                490                495
Gly Asn Cys Asn Glu Glu Ala Ser Gln Ser Cys Pro Asn Glu Lys Cys
                500                505                510
Tyr Leu Ile Leu Asp Cys Ser Gly Phe Thr Phe Phe Asp Tyr Ser Gly
515                520                525
Val Ser Met Leu Val Glu Val Tyr Met Asp Cys Lys Gly Arg Ser Val
530                535                540
Asp Val Leu Leu Ala His Cys Thr Ala Ser Leu Ile Lys Ala Met Thr
545                550                555                560
Tyr Tyr Gly Asn Leu Asp Ser Glu Lys Pro Ile Phe Phe Glu Ser Val
                565                570                575
Ser Ala Ala Ile Ser His Ile His Ser Asn Lys Met Glu Ser Arg Ser
580                585                590
Val Ser His Ala Gly Val Ser Arg Ser Arg Leu Thr Ala Ser Ser Ala
595                600                605
Ser Arg Val His Ala Phe Leu Leu Pro Gln Pro Leu Glu
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<210> 5

<211> 1992

<212> DNA

<213> homo sapiens

<400> 5

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caggtgaccc aaggattggc ctttctgtt ctctcatctg tgcacccagt gtttggttta     240
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acaggcacct ttgccttgac atccttaata tcagccaacg ccgtggaacg gattgtccct     360
cagaacatgc agaatctcac cacacagagt aacacaagcg tgctgggctt atccgacttt     420
gaaatgcaaa ggatccacgt tgctgcagca gtttccttct tgggaggtgt gattcaggtg     480
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gaaaacatca agtctgtgcg actggaagca ttgcttttat ccttctgtgag cattgtggtc     720
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<210> 6

<211> 663

<212> PRT

<213> homo sapiens

<400> 6

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      20           25           30
Pro Ile Leu Asp Trp Ala Pro His Tyr Asn Leu Lys Glu Asn Leu Leu
      35           40           45
Pro Asp Thr Val Ser Gly Ile Met Leu Ala Val Gln Gln Val Thr Gln
      50           55           60
Gly Leu Ala Phe Ala Val Leu Ser Ser Val His Pro Val Phe Gly Leu
      65           70           75           80
Tyr Gly Ser Leu Phe Pro Ala Ile Ile Tyr Ala Ile Phe Gly Met Gly
      85           90           95
His His Val Ala Thr Gly Thr Phe Ala Leu Thr Ser Leu Ile Ser Ala
      100           105           110

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Asn	Ala	Val	Glu	Arg	Ile	Val	Pro	Gln	Asn	Met	Gln	Asn	Leu	Thr	Thr	115	120	125
Gln	Ser	Asn	Thr	Ser	Val	Leu	Gly	Leu	Ser	Asp	Phe	Glu	Met	Gln	Arg	130	135	140
Ile	His	Val	Ala	Ala	Ala	Val	Ser	Phe	Leu	Gly	Gly	Val	Ile	Gln	Val	145	150	155
Ala	Met	Phe	Val	Leu	Gln	Leu	Gly	Ser	Ala	Thr	Phe	Val	Val	Thr	Glu	165	170	175
Pro	Val	Ile	Ser	Ala	Met	Thr	Thr	Gly	Ala	Ala	Thr	His	Val	Val	Thr	180	185	190
Ser	Gln	Val	Lys	Tyr	Leu	Leu	Gly	Met	Lys	Met	Pro	Tyr	Ile	Ser	Gly	195	200	205
Pro	Leu	Gly	Phe	Phe	Tyr	Ile	Tyr	Ala	Tyr	Val	Phe	Glu	Asn	Ile	Lys	210	215	220
Ser	Val	Arg	Leu	Glu	Ala	Leu	Leu	Leu	Ser	Leu	Leu	Ser	Ile	Val	Val	225	230	235
Leu	Val	Leu	Val	Lys	Glu	Leu	Asn	Glu	Gln	Phe	Lys	Arg	Lys	Ile	Lys	245	250	255
Val	Val	Leu	Pro	Val	Asp	Leu	Val	Leu	Ile	Ile	Ala	Ala	Ser	Phe	Ala	260	265	270
Cys	Tyr	Cys	Thr	Asn	Met	Glu	Asn	Thr	Tyr	Gly	Leu	Glu	Val	Val	Gly	275	280	285
His	Ile	Pro	Gln	Gly	Ile	Pro	Ser	Pro	Arg	Ala	Pro	Pro	Met	Asn	Ile	290	295	300
Leu	Ser	Ala	Val	Ile	Thr	Glu	Ala	Phe	Gly	Val	Ala	Leu	Val	Gly	Tyr	305	310	315
Val	Ala	Ser	Leu	Ala	Leu	Ala	Gln	Gly	Ser	Ala	Lys	Lys	Phe	Lys	Tyr	325	330	335
Ser	Ile	Asp	Asp	Asn	Gln	Glu	Phe	Leu	Ala	His	Gly	Leu	Ser	Asn	Ile	340	345	350
Val	Ser	Ser	Phe	Phe	Phe	Cys	Ile	Pro	Ser	Ala	Ala	Ala	Met	Gly	Arg	355	360	365
Thr	Ala	Gly	Leu	Tyr	Ser	Thr	Gly	Ala	Lys	Thr	Gln	Val	Ala	Cys	Leu	370	375	380
Ile	Ser	Cys	Ile	Phe	Val	Leu	Ile	Val	Ile	Tyr	Ala	Ile	Gly	Pro	Leu	385	390	395
Leu	Tyr	Trp	Leu	Pro	Met	Cys	Val	Leu	Ala	Ser	Ile	Ile	Val	Val	Gly	405	410	415
Leu	Lys	Gly	Met	Leu	Ile	Gln	Phe	Arg	Asp	Leu	Lys	Lys	Tyr	Trp	Asn	420	425	430
Val	Asp	Lys	Ile	Asp	Trp	Gly	Ile	Trp	Val	Ser	Thr	Tyr	Val	Phe	Thr	435	440	445
Ile	Cys	Phe	Ala	Ala	Asn	Val	Gly	Leu	Leu	Phe	Gly	Val	Val	Cys	Thr	450	455	460
Ile	Ala	Ile	Val	Ile	Gly	Arg	Phe	Pro	Arg	Ala	Met	Thr	Val	Ser	Ile	465	470	475
Lys	Asn	Met	Lys	Glu	Met	Glu	Phe	Lys	Val	Lys	Thr	Glu	Met	Asp	Ser	485	490	495
Glu	Thr	Leu	Gln	Gln	Val	Lys	Ile	Ile	Ser	Ile	Asn	Asn	Pro	Leu	Val	500	505	510
Phe	Leu	Asn	Ala	Lys	Lys	Phe	Tyr	Thr	Asp	Leu	Met	Asn	Met	Ile	Gln	515	520	525
Lys	Glu	Asn	Ala	Cys	Asn	Gln	Pro	Leu	Asp	Asp	Ile	Ser	Lys	Cys	Glu	530	535	540
Gln	Asn	Thr	Leu	Leu	Asn	Ser	Leu	Ser	Asn	Gly	Asn	Cys	Asn	Glu	Glu	545	550	555

[illegible]

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<210> 7
<211> 1818
<212> DNA
<213> homo sapiens
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<210> 8
<211> 605
<212> PRT
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<213> homo sapiens

<400> 8

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      20          25          30
Pro Ile Leu Asp Trp Ala Pro His Tyr Asn Leu Lys Glu Asn Leu Leu
      35          40          45
Pro Asp Thr Val Ser Gly Ile Met Leu Ala Val Gln Gln Val Thr Gln
      50          55          60
Gly Leu Ala Phe Ala Val Leu Ser Ser Val His Pro Val Phe Gly Leu
65          70          75          80
Tyr Gly Ser Leu Phe Pro Ala Ile Ile Tyr Ala Ile Phe Gly Met Gly
      85          90          95
His His Val Ala Thr Gly Thr Phe Ala Leu Thr Ser Leu Ile Ser Ala
      100          105          110
Asn Ala Val Glu Arg Ile Val Pro Gln Asn Met Gln Asn Leu Thr Thr
      115          120          125
Gln Ser Asn Thr Ser Val Leu Gly Leu Ser Asp Phe Glu Met Gln Arg
      130          135          140
Ile His Val Ala Ala Ala Val Ser Phe Leu Gly Gly Val Ile Gln Val
145          150          155          160
Ala Met Phe Val Leu Gln Leu Gly Ser Ala Thr Phe Val Val Thr Glu
      165          170          175
Pro Val Ile Ser Ala Met Thr Thr Gly Ala Ala Thr His Val Val Thr
      180          185          190
Ser Gln Val Lys Tyr Leu Leu Gly Met Lys Met Pro Tyr Ile Ser Gly
      195          200          205
Pro Leu Gly Phe Phe Tyr Ile Tyr Ala Tyr Val Phe Glu Asn Ile Lys
      210          215          220
Ser Val Arg Leu Glu Ala Leu Leu Leu Ser Leu Leu Ser Ile Val Val
225          230          235          240
Leu Val Leu Val Lys Glu Leu Asn Glu Gln Phe Lys Arg Lys Ile Lys
      245          250          255
Val Val Leu Pro Val Asp Leu Val Leu Ile Ile Ala Ala Ser Phe Ala
      260          265          270
Cys Tyr Cys Thr Asn Met Glu Asn Thr Tyr Gly Leu Glu Val Val Gly
      275          280          285
His Ile Pro Gln Gly Ile Pro Ser Pro Arg Ala Pro Pro Met Asn Ile
      290          295          300
Leu Ser Ala Val Ile Thr Glu Ala Phe Gly Val Ala Leu Val Gly Tyr
305          310          315          320
Val Ala Ser Leu Ala Leu Ala Gln Gly Ser Ala Lys Lys Phe Lys Tyr
      325          330          335
Ser Ile Asp Asp Asn Gln Glu Phe Leu Ala His Gly Leu Ser Asn Ile
      340          345          350
Val Ser Ser Phe Phe Phe Cys Ile Pro Ser Ala Ala Ala Met Gly Arg
      355          360          365
Thr Ala Gly Leu Tyr Ser Thr Gly Ala Lys Thr Gln Val Ala Cys Leu
      370          375          380
Ile Ser Cys Ile Phe Val Leu Ile Val Ile Tyr Ala Ile Gly Pro Leu
385          390          395          400
Leu Tyr Trp Leu Pro Met Cys Val Leu Ala Ser Ile Ile Val Val Gly
      405          410          415
Leu Lys Gly Met Leu Ile Gln Phe Arg Asp Leu Lys Lys Tyr Trp Asn

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450	455	460
Leu Met Asn Met Ile Gln Lys Glu Asn Ala Cys	Asn Gln Pro Leu Asp	
465	470	475
Asp Ile Ser Lys Cys Glu Gln Asn Thr Leu Leu	Asn Ser Leu Ser Asn	
485	490	495
Gly Asn Cys Asn Glu Glu Ala Ser Gln Ser Cys	Pro Asn Glu Lys Cys	
500	505	510
Tyr Leu Ile Leu Asp Cys Ser Gly Phe Thr Phe	Phe Asp Tyr Ser Gly	
515	520	525
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Tyr Gly Ser Leu Phe Pro Ala Ile Ile Tyr Ala Ile Phe Gly Met Gly
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Gln Ser Asn Thr Ser Val Leu Gly Leu Ser Asp Phe Glu Met Gln Arg
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Pro Asp Thr Val Ser Gly Ile Met Leu Ala Val Gln Gln Val Thr Gln
50     55     60
Gly Leu Ala Phe Ala Val Leu Ser Ser Val His Pro Val Phe Gly Leu
65     70     75     80
Tyr Gly Ser Leu Phe Pro Ala Ile Ile Tyr Ala Ile Phe Gly Met Gly
85     90     95
His His Val Ala Thr Gly Thr Phe Ala Leu Thr Ser Leu Ile Ser Ala
100    105    110
Asn Ala Val Glu Arg Ile Val Pro Gln Asn Met Gln Asn Leu Thr Thr
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Gln Ser Asn Thr Ser Val Leu Gly Leu Ser Asp Phe Glu Met Gln Arg
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Pro Val Ile Ser Ala Met Thr Thr Gly Ala Ala Thr His Val Val Thr
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